

A trypanocidal diterpene with novel skeleton from *Dracocephalum komarovi*

Nahoko Uchiyama,^a Michiho Ito,^a Fumiyuki Kiuchi,^b Gisho Honda,^{a,*} Yoshio Takeda,^c
Olimjon K. Khodzhimatov^d and Ozodbek A. Ashurmetov^d

^aGraduate School of Pharmaceutical Sciences, Kyoto University, Kyoto 606-8501, Japan

^bTsukuba Medicinal Plant Research Station, National Institute of Health Sciences, Tsukuba 305-0843, Japan

^cFaculty of Integrated Arts and Sciences, The University of Tokushima, Tokushima 770-8502, Japan

^dScientific Production Center 'Botanika' of Uzbek Academy of Sciences, Tashkent 700-143, Uzbekistan

Received 1 October 2003; revised 30 October 2003; accepted 31 October 2003

Abstract—A new diterpene, komarovspirone (**1**) with a spiro-octahydroindene skeleton, was isolated from *Dracocephalum komarovi*. The structure was elucidated by extensive analyses of spectral data. Komarovspirone (**1**) showed trypanocidal activity against epimastigote of *Trypanosoma cruzi*, the causative agent of American trypanosomiasis, with a minimum lethal concentration of 23 μ M.

© 2003 Elsevier Ltd. All rights reserved.

Dracocephalum komarovi Lipsky (Labiatae) is a perennial semishrub¹ that is called 'buzbosh' in Uzbekistan, and local people use the aerial parts in a tea to treat various diseases such as inflammatory diseases and hypertony. In a previous paper, we reported the isolation of three trypanocidal diterpenes from this plant.² In this paper, we report the isolation and structural determination of a new diterpene with a novel skeleton, komarovspirone (**1**).

Dried whole plants of *D. komarovi* were extracted and fractionated as described previously.² Separation of the fractions with strong trypanocidal activity against epimastigotes of *Trypanosoma cruzi* by silica gel column chromatography (hexane–AcOEt, CHCl₃–acetone, benzene–AcOEt, benzene–acetone) and HPLC (YMC Pack SIL-06, hexane–AcOEt) gave komaroviquinone (**3**) as the major trypanocidal compound.² Separation of a side fraction from the first silica gel column (hexane–AcOEt), which showed moderate trypanocidal activity, by silica gel column chromatography (hexane–acetone, benzene–AcOEt) and HPLC (YMC Pack SIL-06, hex-

ane–AcOEt = 15:1) resulted in isolation of compound **1** (10 mg).³

Compound **1** was obtained as yellow needles from MeOH, $[\alpha]_D^{25} +282.6^\circ$ (*c* 1.0, MeOH). The molecular formula of C₂₁H₂₈O₅ was revealed by high-resolution EIMS (HR-EIMS) (M⁺ *m/z* 360.1924; calculated, 360.1937). The NMR spectra of **1** (Table 1) showed the presence of a methoxy group (δ_C 59.8, δ_H 3.71), a chelated hydroxy group (δ_H 13.32), two singlet methyls (δ_C 31.6, δ_H 0.53 and δ_C 19.8, δ_H 0.87), two methylenes [δ_C 40.3, δ_H 1.94 (1H, t, *J* = 11.9 Hz) and 1.47 (1H, dd, *J* = 11.6, 9.8 Hz); δ_C 43.6, δ_H 2.06 and 1.20, (each 1H, d, *J* = 11.9 and 12.2 Hz)] and an isopropyl group [δ_C 26.0, δ_H 3.57 (1H, sept, *J* = 7.0 Hz); δ_C 20.6, δ_H 1.37 (3H, d, *J* = 7.1 Hz); δ_C 20.4, δ_H 1.36 (3H, d, *J* = 7.0 Hz)]. These spectra were similar to those of the icetexane diterpene, komaroviquinone (**3**),² and the HMBC spectrum revealed that the hydroxy, methoxy and isopropyl groups were in the same arrangement as in **3** (Fig. 1). However, protons of the methylenes corresponding to C-6 (δ_C 40.3) and C-20 (δ_C 43.6) in **1** showed correlation peaks with the carbonyl carbon at δ_C 195.5, the oxygenated quaternary carbon at δ_C 91.5, and a quaternary carbon at δ_C 51.7, indicating the skeleton to be different from that of icetexane in the C-7 to C-9 part. The protons of the two methylenes also showed correlations with the olefinic carbon (δ_C 107.1), which was correlated to the

Keywords: *Dracocephalum komarovi*; Diterpene; Spiro lactone; *Trypanosoma cruzi*; Trypanocidal activity.

* Corresponding author. Tel.: +81-75-753-4524; fax: +81-75-753-4591; e-mail: ghonda@pharm.kyoto-u.ac.jp

Table 1. NMR data of **1** in $C_6D_6^a$

No.	^{13}C	1H	HMBC ^b
1	34.4	α 0.89, td, 13.1, 4.6 β 1.75, br d, 13.1	2 10
2	18.5	α 1.25, m, overlap β 1.89, qt, 13.7, 3.4	
3	40.4	α 0.75, td, 13.7, 4.0 β 1.25, m, overlap	19 1, 5
4	33.5	—	—
5	55.4	1.39, dd, 11.0, 9.4	4, 6, 19
6	40.3	α 1.94, t, 11.9 β 1.47, dd, 11.6, 9.8	5, 8, 9, 10, 11 5, 8, 9, 20
7	169.6	—	—
8	107.1	—	—
9	51.7	—	—
10	91.5	—	—
11	195.5	—	—
12	154.7	—	—
13	143.6	—	—
14	160.4	—	—
15	26.0	3.57, sept, 7.0	12, 13, 14, 16, 17
16	20.6 ^c	1.37, d, 7.1 (3H)	13, 15, 17
17	20.4 ^c	1.36, d, 7.0 (3H)	13, 15, 16
18	31.6	0.53, s (3H)	3, 4, 5, 19
19	19.8	0.87, s (3H)	3, 4, 5, 18
20	43.6	2.06, d, 11.9 1.20, d, 12.2	8, 9, 11 5, 6, 8, 10
OMe	59.8	3.71, s (3H)	12
OH		13.32, s	8, 13, 14

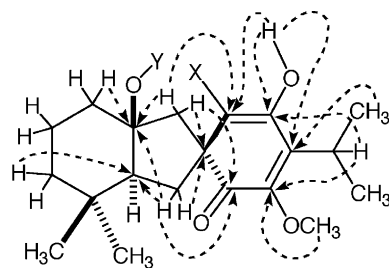
^a 1H NMR, 500 MHz; ^{13}C NMR, 125 MHz; data in δ ppm (J in Hz).

^bCarbons correlated with the proton.

^cThe assignments may be interchanged.

chelated hydroxy proton. From these data and other HMBC correlations, a partial structure as shown in Figure 2 was established. Since one carbon and one oxygen atoms should be added to this partial structure to satisfy its molecular formula $C_{21}H_{28}O_5$, a carbonyl group (δ_C 169.6) was connected to X and Y of the partial structure (Fig. 2) to form a lactone. Thus, the structure of **1** was concluded as indicated, and since it has a novel spiro-octahydroindene skeleton, it is named komarovispirone.

This structure was supported by a chemical conversion. On heating in 5% HCl/MeOH, **1** gave compound **2**.⁴ From HR-EIMS (M^+ m/z 302.1886, calculated, 302.1882), the molecular formula of **2** was revealed to be $C_{19}H_{26}O_3$, which corresponds to simultaneous occurrence of decarboxylation and demethylation. The NMR spectra (Table 2) showed the presence of a hydroxy group (δ_H 7.17), three isolated methylenes (δ_C 42.5, δ_H

**Figure 2.** Selected HMBC correlations and partial structure of **1**.**Table 2.** NMR data of **2** in $CDCl_3^a$

No.	^{13}C	1H	HMBC ^b
1	25.8	1.84, br s (2H)	2, 3, 5, 10
2	19.7	1.62, m (2H)	1, 3, 4, 10
3	38.5	1.40, m (2H)	1, 18, 19
4	32.1	—	—
5	138.9	—	—
6 ^c	47.4	2.85, d, 15.8 2.07, d, 15.8	5, 8, 9, 10 5, 8, 9, 10
8	50.3	2.77, d, 15.5 2.74, d, 15.5	6, 9, 11, 14, 20 6, 9, 11, 14, 20
9	50.1	—	—
10	130.2	—	—
11	198.8	—	—
12	153.5	—	—
13	131.5	—	—
14	196.6	—	—
15	24.7	3.18, sept, 7.2	12, 13, 14, 16, 17
16	19.5	1.20 ^c , d, 7.2 (3H)	13, 15, 17
17	19.5	1.21 ^c , d, 6.9 (3H)	13, 15, 16
18,19	27.5	0.94, s (6H)	3, 4, 5, 18, 19
20 ^c	42.5	2.90, br d, 15.8 2.13, dd, 15.8, 2.1	5, 9, 10 11, 12, 13
OH		7.17, s	11, 12, 13

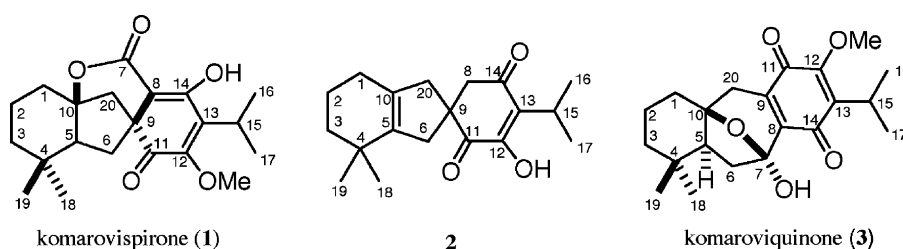
^a 1H NMR, 600 MHz; ^{13}C NMR, 150 MHz; data in δ ppm (J in Hz).

^bCarbons correlated with the proton.

^cThe assignments may be interchanged.

2.90 and 2.13; δ_C 47.4, δ_H 2.85 and 2.07; δ_C 50.3, δ_H 2.77 and 2.74), and two ketone carbonyls (δ_C 196.6 and 198.8), but no signal of the methoxy and lactone groups was observed. Based on the HMBC correlations (Table 2), the structure of **2** was concluded as indicated, and this structure is fully compatible with the structure of **1**.

NOE difference experiments in $CDCl_3$ and/or benzene- d_6 showed NOE effects indicated in Figure 3. Although these results suggested the stereochemistry shown in Figure 3, the entire stereochemistry could not be determined. Biogenetically, komarovispirone (**1**) may be

**Figure 1.**

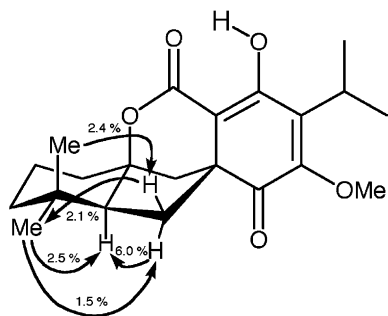
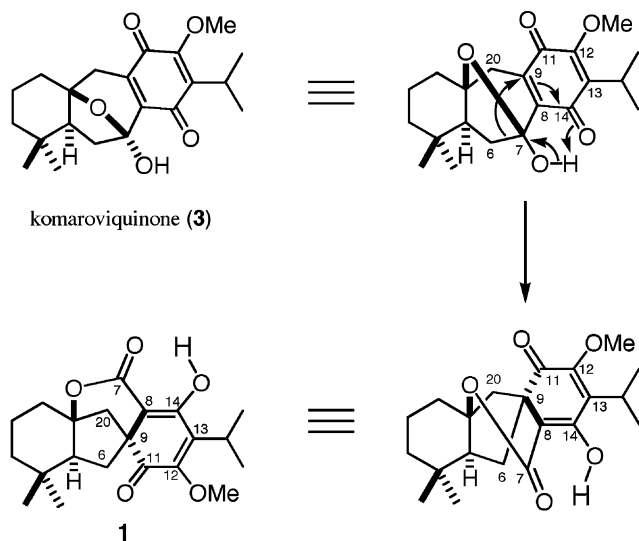


Figure 3. Selected NOEs in **1**.



Scheme 1. Possible route of formation of **1**.

derived from komaroviquinone (**3**), which is isolated from the same plant,² through a rearrangement shown in Scheme 1. Thus, the stereochemistry of **1** was tentatively assigned as indicated.

Komarovispirone (**1**) showed moderate trypanocidal activity against epimastigote of *T. cruzi*⁵ with a minimum lethal concentration (MLC) of 23 μM . The MLC

of **1** was higher than that of **3** (0.4 μM) under the same assay condition.² Several types of natural quinones have been reported to show trypanocidal activity, and their activities have been partly ascribed to production of a reactive oxygen species in the parasite.^{6,7} Thus, the action mechanism of **1** may be similar to that of these quinone compounds.

Acknowledgements

This work was supported in part by a Grant-in-Aid for Scientific Research (No. 12576027) from the Japan Society for the Promotion of Science. We are grateful to Dr. N. Kawahara of National Institute of Health Sciences for 600 MHz NMR measurements and to Dr. N. Akimoto of Kyoto University for MS measurements.

References and Notes

- Vvedenski, A. I. In *Flora Uzbekistana*; Editio Academiae Scientiarum UzSSR: Tashkent, 1961; Vol. 5, p 313.
- Uchiyama, N.; Kiuchi, F.; Ito, M.; Honda, G.; Takeda, Y.; Khodzhimatov, O. K.; Ashurmetov, O. A. *J. Nat. Prod.* **2003**, *66*, 128–131.
- Komarovispirone (**1**): mp 119–120 °C; UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 251 (4.52), 365 (3.06); IR (KBr) cm^{-1} : 2943, 2874, 1651, 1558; EI-MS m/z (%): 360 (M^+ , 26), 316 (100), 301 (61), 283 (17). ^1H NMR (500 MHz) and ^{13}C NMR (125 MHz) in C_6D_6 : see Table 1.
- Komarovispirone **1** (1 mg) was heated in 5% HCl/MeOH (0.5 mL) at 65 °C for 3.5 h. The reaction mixture was concentrated to dryness to give compound **2** (0.6 mg). Compound **2**: colorless amorphous powder, mp > 300 °C; UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 288 (3.95), 212 (3.83); IR (KBr) cm^{-1} : 3360, 2928, 2870, 1666; EI-MS m/z (%): 302 (M^+ , 100), 287 (88). ^1H NMR (600 MHz) and ^{13}C NMR (150 MHz) in CDCl_3 : see Table 2.
- Asaruddin, M. R.; Kiuchi, F.; Honda, G. *Nat. Med.* **2001**, *55*, 149–151.
- Sepúlveda-Boza, S.; Cassels, B. K. *Planta Med.* **1996**, *62*, 98–105.
- Fernandez Villamil, S. H.; Perissinotti, L. J.; Stoppani, O. M. *Biochem. Pharm.* **1996**, *52*, 1875–1882.